Amendments to the Specification:

Please replace the paragraph on page 20, line 32 to page 21, line 9 with the following amended paragraph:

Elastomeric polymers useful as optional secondary polymers in the invention are typically materials that form one phase at 21°C, have a glass transition temperature less than 0°C, and exhibit elastomeric properties. The elastomeric polymers include, but are not limited to, polyisoprenes, styrene-diene block copolymers, natural rubber, polyurethanes, polyether-block-amides, poly-alpha-olefins, (C1-C20) acrylic esters of ~~meth(acrylic)~~ (meth)acrylic acid, ethylene-octene copolymers, and combinations thereof.

Please replace the paragraph on page 20, line 32 to page 21, line 9 with the following amended paragraph:

In some embodiments, a composition containing a silver oxide can be coated on the polymer composition as described in applicants co-pending application, Serial No. 10/728,446, filed December 5, 2003 ~~Ser. No. _____, Attorney Docket No. 59427US002~~, incorporated herein by reference. The metal oxide is dissolved in solution by complexing the metal compound in an ammonium salt. Suitable ammonium salts include ammonium pentaborate, ammonium acetate, ammonium carbonate, ammonium peroxyborate, ammonium tertraborate, triammonium citrate, ammonium carbamate, ammonium bicarbonate, ammonium malate, ammonium nitrate, ammonium nitrite, ammonium succinate, ammonium sulfate, ammonium tartarate, and mixtures thereof. The resultant solution can be coated at less than 40 °C, and dried at temperatures less than 160 °C. Once dried, the substrate remains coated with the metal oxide.

Please replace the line on page 25, line 15 with the following amended line:

Antimicrobial ~~Anti-microbial~~ Performance Tests

Please replace the paragraph on page 25, line 32 to page 26, line 14 with the following amended paragraph:

Bacteria labeling and Antimicrobial ~~Anti-microbial~~ testing: 7 mls of bacteria solution at initial concentration of approximately $1x10^8$ bacteria/mls were pipetted into

a 50 mls conical tube containing the sample. At the specified time (e.g., 2 hr), 50 micro-liter (μL) of the supernatant was pipetted into fluorescent measurement tube which already contained 450 μL of MiliQ water and premixed green dye and red dye solution (1.5 μL dye mixture for 500 μL bacteria solution) was added and the mixture was incubated for 15 minutes in the dark at room temperature. These solutions were then measured by flow cytometry. Cell viability was measured using the BD FACS Caliber flow cytometer (made by Becton Dickinson & Company, Franklin Lakes, New Jersey). The flow cytometer is equipped with an argon-ion laser at 488 nanometers (nm) and 15 milliWatts (mW) output. Data acquisition and analysis were controlled using CellQuest software and PBPAC hardware interface. The light path contained a 488/10 nm blocking filter, then a 530/30 nm filter before the green PMT and a 585/42 nm long pass filter before the red PMT. The sampling rate was around 3000-7000 particles/second. The sheath fluid was FACSFlow by Becton Dickinson. The instrument voltage was 5.5 Volt.

Please replace the paragraph on page 26, line 25 to page 27, line 3 with the following amended paragraph:

Antimicrobial ~~Anti-microbial~~ performance was measured using a Zone of Inhibition test (ZOI) that was performed by the following method. Mueller-Hinton agar was prepared, sterilized and tempered in a water bath at 48-50°C. A suspension of bacteria in sterile phosphate-buffered water was prepared with approximately 10^8 CFU/ml. The agar was inoculated with the bacterial suspension to an approximate concentration of 10^5 CFU/ml (1:1000). The inoculated agar was swirled to mix and pipetted (~14 ml) into sterile Petri dishes (15 x 100 mm). The seeded agar was allowed to set for about 20 minutes to harden. An alcohol-disinfected die and cutting board were used to cut textile samples to desired size. Sterile forceps were used to place the samples onto the seeded, hardened agar in center of plate. The plate was then placed into an incubator at 35-37°C for overnight (16-24 hours) incubation. After incubation the clear zones, no visible colonies formed, were measured in mm with calipers.

Please replace the paragraph on page 32, lines 18-24 with the following amended paragraph:

Example 3 was tested for antimicrobial ~~anti-microbial~~ performance against *Staph. Aureus* using the Zone of Inhibition Test. Example 3 was sterilized using a cobalt-γ source at both 25 and 40 kilograys (kGy). The samples were tested in the dry state. All samples had a diameter of 24 mm. Table 7 contains the results from the Zone of Inhibition Test for Example 3 at two sterilization exposure levels and a commercially available silver dressing, Example 5 (Comparative-ACTICOAT available from Smith and Nephew, Largo, Florida).

Please replace the paragraph on page 34, starting after "Table 9" to page 35, line 3 with the following amended paragraph:

Comparative Example 6 and Examples 7-10 were tested for antimicrobial ~~anti-microbial~~ activity against *Staph. aureas* using the % Live Bacteria Test. One drop of the Example dispersions was dripped into the bacterial solution and mixed. The % live bacteria at 2 hours was measured. All bacterial solution volumes were 7 mL. The initial live bacteria concentration was 1.0×10^8 bacteria/mL. The results are tabulated in Table 10.